

SYNTHESIS OF 7-*O*-[2,6-DIDEOXY-2-FLUORO-4-*O*-(3-FLUOROTETRAHYDROPYRAN-2-YL)- α -L-TALOPYRANOSYL]DAUNOMYCINONEYASUSHI TAKAGI, HIROMI SOHTOME, TSUTOMU TSUCHIYA*
and SUMIO UMEZAWAInstitute of Bioorganic Chemistry,
1614 Ida, Nakahara-ku,
Kawasaki 211, Japan

TOMIO TAKEUCHI

Institute of Microbial Chemistry,
3-14-23 Kamiosaki,
Shinagawa-ku, Tokyo 141, Japan

(Received for publication August 23, 1991)

7-*O*-[2,6-Dideoxy-2-fluoro-4-*O*-(3-fluorotetrahydropyran-2-yl)- α -L-talopyranosyl]daunomycinones (**7a**~**7c**) have been prepared by condensation of 3'-*O*-benzoyl derivative of 7-*O*-(2,6-dideoxy-2-fluoro- α -L-talopyranosyl)daunomycinone with 2,3-difluorotetrahydropyran as the key reaction. Antitumor activities of these compounds are described.

Recently we reported the synthesis and antitumor activities of 7-*O*-(2,6-dideoxy-2-fluoro- α -L-talopyranosyl)daunomycinone (**1**) and -adriamycinone^{1,2)} (**2**) together with their analogs.^{3~5)} Most of them showed higher antitumor activities than doxorubicin and daunorubicin, and showed much lower toxicity. Characteristic features of these compounds in terms of chemical structure are that some of them have no amino group at C-3',^{1~3,5)} which has been believed to be essential for activity, and most of the compounds have a C-2' fluorine,^{1~4)} no functional group being attached here in the usual anthracycline antitumor antibiotics. On the other hand, structurally related aclacinomycins^{6,7)} and (2'*R*)-4'-*O*-tetrahydropyranyladriamycin,^{8,9)} have a 4'-*O*-glycosyl moiety and exhibit lower toxicity with higher antitumor activity^{10,11)} than doxorubicin. Further, the last compound was reported¹²⁾ to be readily taken up by tumor cells. With all of these facts in mind, we have undertaken to prepare several 4'-*O*-glycosyl derivatives of **1** and **2**. This paper describes the synthesis and antitumor activity of 4'-*O*-(3-fluorotetrahydropyran-2-yl) derivatives (**7**) of **1**. As these compounds **7** have a tetrahydropyran moiety bearing a fluorine at the vicinal position to the glycosyl bond, the moiety would be expected to resist acid-catalyzed hydrolysis due to the strong electron-withdrawing property of the vicinal fluorine; in the cases of aclacinomycins and 4'-*O*-tetrahydropyranyl derivatives, the 4'-*O*-glycosyl group is readily cleaved chemically. We undertook the preparation of **7** to test whether the enhanced chemical stability would also be retained in *in vivo* experiments.

Treatment of **1** with benzoyl chloride in dichloromethane - pyridine gave selectively the 3'-*O*-benzoyl derivative (**3**) with the axial 4'-OH remained free. Next, in order to introduce 3-fluorotetrahydropyran-2-yl group at the 4'-OH, 2,3-difluorotetrahydropyran was first prepared. Introduction of fluorine gas into 3,4-dihydro-2*H*-pyran in cold (-78°C) dichloromethane readily gave the difluoro derivatives (**4** and **5**); however, these products were markedly unstable in an acidic medium (in this case, HF was formed inevitably), and decomposed on concentration, so that they could not be isolated in a pure state. Therefore, the above reaction mixture was immediately coupled with **3** in the presence of AgClO₄, SnCl₂, and molecular

Fig. 1. Structures of 1~3 and 4'-O-(3-fluorotetrahydropranyl) derivatives of 1.

	R ₁	R ₂	R ₃
1	H	H	H
2	OH	H	H
3	H	C ₆ H ₅ CO	H
6	H	C ₆ H ₅ CO	
7a	H	H	
7b	H	H	
7c	H	H	

Table 1. Coupling constants (Hz) of the ¹⁹F NMR spectra for 4 and 5 in CD₂Cl₂.

	4	5
$J_{2,2-F}$	55	50
$J_{3,2-F}$	24	≤2
$J_{2,3-F}$	≤2	~4
$J_{3,3-F}$	47	47
$J_{4a,3-F}$	4~5	58
$J_{4b,3-F}$	4~5	~4
$J_{2-F,3-F}$	19.5	15
One wing of the signal range of 3-F	50	80

sieves in dichloromethane to give a mixture of 4'-O-(3-fluorotetrahydropranyl-2-yl) derivatives (6) in good yield. The structures of the 2,3-difluorotetrahydropranyl, however, could be proved by the ¹H and ¹⁹F NMR spectra on the sample prepared as follows: After treatment of 3,4-dihydro-2H-pyran with F₂ in argon in cold (-78°C) dichloromethane-

*d*₂, the solution was washed thoroughly with cold aqueous sodium thiosulfate, and, without delay, the ¹H and ¹⁹F NMR spectra were measured; the ¹⁹F spectrum showed clear signals (Table 1) for *cis* (4) and *trans* 2,3-difluorotetrahydropranyl in a 7.5:2.5 ratio. The coupling constants indicate (based on the reported coupling constants¹³) for 2-fluoroglycopyranosyl fluorides) that both 4 and 5 have a chair conformation, respectively, having 2-axial-3-equatorial (for 4) and 2,3-diaxial fluorines (for 5). It was surprising that these fundamental substances 4 and 5 have not been reported previously.

Debenzoylation of 6 with methanolic sodium hydroxide gave the three final products (7a, 7b, and 7c). Separation of them was, however, tedious and performed only by repeated column chromatography and HPLC to give pure products in 31.4 (7a), 6.4 (7b), and 20.1% (7c), respectively. The structures of the products were determined from their NMR spectra (¹H, ¹⁹F, and ¹³C (Table 2)) to be the expected 4'-O-(3-fluorotetrahydropranyl-2-yl) derivatives of 1, but the absolute configurations at C-2'' and C-3'' remained undetermined. However, judging from the ¹H and ¹⁹F NMR data as shown in Table 3 (in some signal-regions assignments are difficult due to bad signal dissolution or overlapping), the structures could partly be clarified. First, compound 7b is determined to be a 2'',3''-*cis* product (the large $J_{3'',4''a}$ value, and narrow signal-range of one wing of the broad 3''-F doublet suggest the equatorial 3''-F, and the small $J_{2'',3''}$ value suggests the axial 2''-OR) with a chair conformation of 2''C₅ (3''S) (C) or 5''C₂ (3''R) (C') (see Fig. 2). Compound 7a was concluded to have a 2'',3''-*trans* structure with 3''S or 3''R as a mixture of two chair conformations (A and B, or D and E) (judged from relatively large $J_{2'',3''}$ value (6 Hz) and relatively large signal-range of one wing of the broad 3''-F doublet (Table 3)). It should be stressed here

Table 2. ^{13}C NMR data of compounds **7a**, **7b**, and **7c** (in CDCl_3).

C	7a	7b	7c
1	119.7	119.9	119.8
2	135.7	135.8	135.7
3	118.5	118.6	118.5
4	161.1	161.2	161.1
4a	120.8	121.0	120.8
5	186.8 ^a	187.1 ^a	186.9 ^a
5a	111.5 ^b	111.65 ^b	111.5 ^b
6	156.2 ^c	156.3 ^c	156.3 ^c
6a	134.2 ^d	134.3 ^d	134.2 ^d
7	70.9	70.8	70.8
8	35.1	35.2	35.1
9	76.5	^g	76.5
10	33.1	33.3	33.2
10a	135.4 ^d	135.6 ^d	135.4 ^d
11	155.5 ^c	155.7 ^c	155.5 ^c
11a	111.4 ^b	111.59 ^b	111.4 ^b
12	186.7 ^a	186.9 ^a	186.7 ^a
12a	133.3 ^d	133.4 ^d	133.3 ^d
13	211.3	211.4	211.3
14	24.6	24.6	24.6
OMe	56.6	56.7	56.7
1'	100.9 and 101.5, d	101.2 and 101.8, d	101.3 and 101.8, d
2'	85.8 and 88.6, d	86.3 and 89.1, d	86.1 and 89.0 ^e , d
3'	66.5 and 66.7, d	66.3 and 66.5, d	65.7 and 66.0, d
4'	78.5	82.3	82.3
5'	67.0	67.7	67.4
6'	16.9	16.5	16.4
2''	102.7 and 103.1, d	99.4 and 99.8, d	102.7 and 103.1, d
3''	87.3 and 90.1, d	86.5 and 89.4, d	86.3 and 89.2 ^e , d
4''	27.4 and 27.7, d	24.0 and 24.3, d	26.6 and 26.9, d
5''	23.0 and 23.1, d	23.6 and 23.8, d	22.1 and 22.2, d
6''	64.2	60.8 and 60.9, d	64.2
<i>J</i> (Hz)			
<i>J</i> _{1'-C,2'-F}	32.4	32.3	32.3
<i>J</i> _{2'-C,2'-F}	177.9	179.3	180.3 ^f
<i>J</i> _{3'-C,2'-F}	16.9	17.2	16.9
<i>J</i> _{2''-C,3''-F}	25.0	22.4	27.3
<i>J</i> _{3''-C,3''-F}	175.9	184.0	177.5 ^f
<i>J</i> _{4''-C,3''-F}	19.0	18.9	19.4
<i>J</i> _{5''-C,3''-F}	7.0	8.4	6.2
<i>J</i> _{6''-C,3''-F}		7.0	

^{a,b,c,d,e,f} Figures in the same column may be interconvertible.

^g Overlapped with the signal of CDCl_3 .

that the observed coupling ($J=6$ Hz) between 3'-OH and 3''-F is considered to be a through-space coupling. This indicates that both atoms come close to each other, suggesting the contribution of the A conformation of **7a** (see later in detail). Compound **7c** will be a 2'',3''-*trans* product, judging from the similarity of the signal pattern with that for **7a** and therefore, it will be an enantiomer of **7a** in terms of its 3-fluorotetrahydropyranyl portion, and the conformation will be a mixture of D and E, or A and B. ^1H -Shifts of 2''-H for **7a**, **7b**, and **7c** also support the above conclusions, that is, 2''-H of **7b** is the most deshielded (Table 3), whereas the 2''-H's of **7a** and **7c** are comparatively shielded suggesting the involvement of the axial 2''-H.

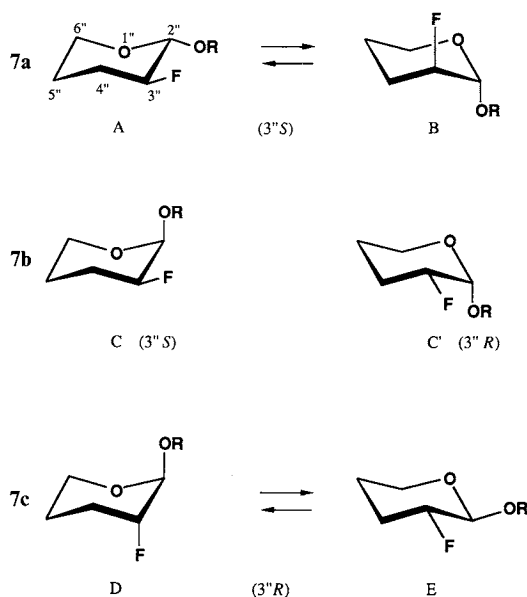
Table 3. ^1H and ^{19}F NMR data relating to the 3''-F and optical rotations of **7a**, **7b**, and **7c**.

	7a	7b	7c
2''-H (δ)	4.66 (dd)	4.83 (t)	4.45 (t)
3''-H (δ)	4.45	4.56	4.44
Signal range for 3''-H (Hz)	20	23	20
3''-F (δ)	-186.2 (br d)	-186.0 (br d)	-187.3 (dddd) ^a
Signal range of one wing of 3''-F (Hz)	47	33	50
$J_{2'',3''}$ (Hz)	6	3.5	5
$J_{3'',4''a}$		~11	
$J_{3'',4''b}$		~5	
$J_{2'',F}$	5	3	5
$J_{3'',F}$	~50	~50	50
$J_{4''a,F}$			~21
$J_{4''b,F}$			~12
$[\alpha]_D$ (c 0.1, CHCl_3)	+148°	+55°	+101°

^a Slightly deformed dddd.

Although, as described above, absolute configurations of the 3-fluorotetrahydropyranyl portion at C-2'' and C-3'' of **7a**, **7b**, and **7c** remained undetermined, the large difference in optical rotations among them (Table 3) suggests their configurations. In simple alkyl glycohexopyranosides with an equatorial C-5 substituent (the configuration at C-5 determines D and L), four types of chair conformations (I, II, III, and IV; see Fig. 3) exist and their optical rotations are generally determined by the types ($\alpha\text{-D} > \beta\text{-L} > \beta\text{-D} > \alpha\text{-L}$). This phenomenon originates from the difference in electron-density around the anomeric center. If such an experimental rule is applied to our cases (although the C-6'' substituents of **7a**~**7c** are two hydrogens, there will be little difference from the alkyl substituent in I~IV in terms of the contribution to the rotations), the following assignments will be made. As the contribution of the 3-fluorotetrahydropyranyl portion of **7b** to its rotation is expected to be highly negative, structure C (resembles IV), rather than C' is assigned. Compound **7a** has the highest rotational value, and therefore it should be a mixture of A and B (resembles II and I) rather than that of D and E (resembles IV and III) and **7c**, *vice versa*. In conclusion, we decided that the structures of **7a**, **7b**, and **7c** are as shown in Fig. 1.

Antitumor activities of **7a**, **7b**, and **7c** were measured (Tables 4 and 5) and it was concluded that the three compounds and **1** have similar activity. However, as the three compounds were non-toxic in the amount of 100 $\mu\text{g}/\text{mouse}/\text{day}$ (Table 5), this derivation will give less toxic derivatives compared to the parent

Fig. 2. Conformations of **7a**, **7b**, and **7c**.

R = [7-O-(2,6-Dideoxy-2-fluoro- α -L-talopyranosyl)-daunomycinone]-4'-yl

anthracycline antibiotics. Other biological characteristics will be reported in the future with other related compounds.

Experimental

Optical rotations were measured with a Perkin-Elmer 241 polarimeter. TLC was carried out on precoated Kieselgel 60 F₂₅₄ (Merck). Column chromatography was performed on Wakogel C-200. HPLC was performed with a SSC-6200 instrument (Senshu Scientific Co.) with a UV detector (SSC 3000A-II) set at 290 nm. NMR spectra (¹H at 250 MHz, ¹⁹F at 235.3 MHz, and ¹³C at 62.9 MHz) were recorded with a Bruker WM 250 spectrometer. Chemical shifts (δ) are reported downfield from internal tetramethylsilane or Freon 11 (CFCl₃; for ¹⁹F).

7-O-(3-O-Benzoyl-2,6-dideoxy-2-fluoro- α -L-talopyranosyl)daunomycinone (3)

To a suspension of **1** (72.6 mg, 0.13 mmol) in pyridine-dichloromethane (1:8, 1.4 ml) was added a solution of benzoyl chloride (16.2 μ l, 0.14 mmol) in dichloromethane (0.32 ml), and the mixture was stirred at room temperature for 2 hours. The resulting solution showed, on TLC with C₆H₆-Me₂CO (4:1), spots at R_f 0.4 (**3**, major) and 0.7 (trace). Methanol (10 μ l) and then CHCl₃ (20 ml) were added and the solution was washed successively with 20% aq KHSO₄, saturated aq NaHCO₃, and water, dried (Na₂SO₄), and concentrated. The residue was separated by silica gel column chromatography with CHCl₃-Me₂CO (12:1) to give **3** as a reddish-orange solid, 78 mg (90%). An analytical sample was obtained by reprecipitation from CHCl₃-hexane: $[\alpha]_D^{25} +168^\circ$ (c 0.1, CHCl₃); ¹H NMR (CDCl₃) δ 1.41 (3H, d, *J*_{5,6} = 6.5 Hz,

Table 4. Growth inhibitory effect of **7a**, **7b**, and **7c** in comparison with **1** and doxorubicin on P388/S and P388/ADM cells *in vitro*.^a

Compound	IC ₅₀ (ng/ml)		RF ^b
	P388/S	P388/ADM	
7a	13	210	16
7b	33	210	6.4
7c	22	160	7.3
1	33	150	4.5
Doxorubicin	19	920	48

^a IC₅₀ values were determined on day-3 culture.

^b Resistant factor (RF): IC₅₀ for resistant subline against IC₅₀ for sensitive subline.

Fig. 3. Relationship between optical rotations and glycohexopyranoside structures with an equatorial C-5 substituent.

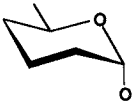
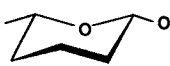
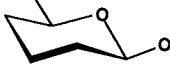
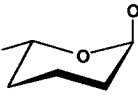
				
	I	II	III	IV
Optical rotation	α -D ++	β -L + or ~0	β -D - or ~0	α -L --

Table 5. Antitumor activities (T/C, %) of **7a**, **7b**, and **7c** on L1210.

Compound	Dose (μ g/mouse/day)					
	100	50	25	12.5	6.25	3.13
7a	176	113	110	101	104	101
7b	236	143	107	104	99	101
7c	236	131	113	104	104	104
Daunorubicin ⁴⁾	117*	151*	193	166	133	130

Leukemia L1210 cells (10⁵) were inoculated into CDF₁ mice (20 \pm 1 g) intraperitoneally. Drugs were administered daily, starting 24 hours after inoculation, from day-1 to -9, intraperitoneally.

* Toxic.

5'-Me), 2.21 (1H, dd, $J_{7,8ax}=4.5$, $J_{8ax,8eq}=15$ Hz, 8-H_{ax}), 2.33 (1H, t, $J_{OH,4'}=10.5$, $J_{OH,F} \sim 10$ Hz, 4'-OH), 2.40 (1H, br d, 8-H_{eq}), 2.42 (3H, s, 13-Me), 2.84 (1H, d, $J_{10ax,10eq}=19$ Hz, 10-H_{ax}), 3.17 (1H, dd, $J_{8eq,10eq} \sim 1.5$ Hz, 10-H_{eq}), 3.90 (1H, s, 9-OH), 3.94 (1H, br d, 4'-H), 4.04 (3H, s, OMe), 4.31 (1H, br q, 5'-H), 4.84 (1H, br d, $J_{2',F}=50$ Hz, 2'-H), 5.10 (1H, dt, $J_{2',3'}=J_{3',4'} \sim 3$, $J_{3',F}=33.5$ Hz, 3'-H), 5.29 (1H, br d, 7-H), 5.67 (1H, dd, $J_{1',2'} \sim 1.5$, $J_{1',F}=10$ Hz, 1'-H), 7.35 (1H, br d, $J_{2,3} \sim 8.5$ Hz, 3-H), 7.41 (2H, m, Bz), 7.54 (1H, m, Bz), 7.74 (1H, t, 2-H), 7.95 (1H, br d, $J_{1,2} \sim 7.5$ Hz, 1-H), 8.04 (2H, m, Bz), 13.12 and 13.96 (each 1H, s, 6, 11-OH); ¹⁹F NMR (CDCl₃) δ -198.3 (ddt, $J=50, 33.5, 10$, and 10 Hz).

Anal Calcd for C₃₄H₃₁FO₁₂·H₂O: C 61.08, H 4.97, F 2.84.

Found: C 61.18, H 4.74, F 2.70.

Preparation of DL-2,3-cis-2,3-Difluorotetrahydropyran (4) and DL-2,3-trans-2,3-Difluorotetrahydropyran (5) in CD₂Cl₂

To a cold (-78°C) solution of 3,4-dihydro-2H-pyran (50 mg) in CD₂Cl₂ (2 ml, dried over molecular sieves 4A) was introduced fluorine (600 ml of 5% F₂ in argon) by gentle bubbling for 30 minutes. Nitrogen was then bubbled for a while at the temperature to remove a part of excess fluorine remained. The resulting solution, which is strongly acidic when examined by pH-test paper, was thoroughly washed with ice-cold aqueous Na₂S₂O₃ (5 ml \times 6; washing should be done until the clear aqueous layer was resulted, the turbidity being caused by the sulfur liberated by the reaction between F₂ and this reagent; insufficient washing caused decomposition of the products during storage), aqueous saturated NaHCO₃, aqueous saturated NaCl, and finally with D₂O, and dried over MgSO₄. The solution was directly measured by NMR spectroscopy.

¹H NMR δ 1.4~2.3 (4H, m, 4-H_a, H_b, 5-H_a, H_b), 3.5~4.0 (2H, m, 6-H_a, H_b), 4.3~4.7 (1H, m, 3-H), 5.50 (~0.3H, br d, $J_{2,2-F}=50$ Hz, 2-H of 5), 5.61 (~0.7H, br d, $J_{2,2-F}=55$ Hz, 2-H of 4); ¹⁹F NMR δ -194.5 (~0.26 F, slightly unresolved dddd, 3-F of 5), -189.7 (~0.74 F, unresolved dddd, 3-F of 4), -155.9 (~0.73 F, clear-cut ddd, 2-F of 4), and -142.5 (~0.27 F, dd, 2-F of 5).

7-O-[3-O-Benzoyl-2,6-dideoxy-2-fluoro-4-O-(3-fluorotetrahydropyran-2-yl)- α -L-talopyranosyl]daunomycinone (6)

A stream of 5% F₂ in argon (1.6 liters) was introduced into a solution of 3,4-dihydro-2H-pyran (0.2 g) in dry dichloromethane (4 ml) by gentle bubbling at -78°C for 1 hour. A portion of the resulting solution (2.2 ml) was added immediately to an ice-cold suspension of 3 (166 mg), AgClO₄ (116 mg), SnCl₂ (97 mg), and powdered molecular sieves 4A (830 mg) in dry dichloromethane (3.3 ml), and the mixture was stirred vigorously at 0°C for 4 hours. The supernatant layer showed, on TLC with CHCl₃-Me₂CO (30:1), spots at R_f 0.3 (slight), 0.23 (major, 6), 0.17 (slight), and 0.1 (minor, 3). The reaction mixture was filtered with aid of Celite and the mass was washed thoroughly with CHCl₃. The combined filtrates were washed successively with saturated aq NaHCO₃ and water, dried (Na₂SO₄), and concentrated. The residue was chromatographed on a column of silica gel with CHCl₃-Me₂CO (30:1) to give a red solid of 6 as a mixture of three diastereomers (1:2.6:3.1, judged by ¹⁹F NMR), 152 mg (79%). An analytical sample was prepared by reprecipitation from CHCl₃-hexane. ¹H NMR (CDCl₃) δ 1.36, 1.42, and 1.47 (3H in total, each d, $J_{5',6'}=6.5$ Hz, 5'-Me), 2.43 (3H, s, 13-Me), 4.07 and 4.08 (3H in total, each s, OMe), 13.26 and 13.27 (1H in total, each s, 11- or 6-OH), 14.01, 14.03, and 14.04 (1H in total, each s, 6- or 11-OH); ¹⁹F NMR (CDCl₃) δ -201.8 (0.45 F, ddd, $J_{1'-H,2'-F}=10$, $J_{2'-H,2'-F}=49.5$, $J_{3'-H,2'-F}=32$ Hz, 2'-F), -200.83 (0.15 F, ddd, $J=49.5, 33$, and 10 Hz, 2'-F), -200.82 (0.4 F, ddd, $J=49.5, 33$, and 10 Hz, 2'-F), -189.9 (0.45 F, m, 3''-F), -188.9 (0.4 F, br dddd, $J \sim 49, \sim 37, \sim 12$, and ~ 6.5 Hz, 3''-F), -183.2 (0.15 F, br d, $J \sim 49$ Hz, 3'-F).

Anal Calcd for C₃₉H₃₈F₂O₁₃: C 62.23, H 5.09, F 5.05.

Found: C 62.08, H 5.10, F 4.98.

7-O-[2,6-Dideoxy-2-fluoro-4-O-(3-fluorotetrahydropyran-2-yl)- α -L-talopyranosyl]daunomycinone (7a, 7b, and 7c)

To an ice-cold solution of 6 (196 mg) in oxolane-MeOH (1:1, 20 ml) was added 0.4 M methanolic NaOH (10 ml) and the mixture was stirred at 0°C for 1 hour. The resulting deep-purple solution showed, on TLC with CHCl₃-Me₂CO (15:1), spots at R_f 0.13 (7a) and 0.19 (7b and 7c). After gradual neutralization

with cold aq 0.2M HCl, the mixture was extracted with CHCl_3 . The extract was washed with water, dried (Na_2SO_4), and concentrated. The residue was chromatographed on a column of silica gel with CHCl_3 - Me_2CO (15:1) to give a red solid of **7a**, 36.7 mg, a mixture of **7b** and **7c**, 51.4 mg, and a mixture of **7a**, **7b**, and **7c**, 80 mg. Repeated chromatography of the last mixture with the same solvent system as above afforded an additional crop of **7a**, 16.4 mg, a mixture of **7b** and **7c**, 13 mg, and a mixture of the three, 26.7 mg. Total yield of **7a** was 53.1 mg (31.4%). The combined mixtures of **7b** and **7c** (64.4 mg) were separated by HPLC on a column (30 mm \times 25 cm) of Senshu Pak. SSC-Silica-842, with toluene-MeOH (20:1), at a flow rate of 10 ml/minute, and a pressure of 150 kg/cm², to give a red solid of **7b** (Rt 18.6 minutes), 10.8 mg (6.4%), and **7c** (Rt 20.2 minutes), 34 mg (20.1%). Analytical samples were prepared by reprecipitation from CHCl_3 -hexane.

7a: ¹H NMR (CDCl_3) δ 1.41 (3H, d, $J_{5',6'}=6.5$ Hz, 5'-Me), 1.58~1.83 (3H, m, 4''-H_a, 5''-H_a, H_b), 2.17 (1H, dd, $J_{7,8\text{ax}}=4.5$, $J_{8\text{ax},8\text{eq}}=15$ Hz, 8-H_{ax}), ~2.2 (1H, m, 4''-H_b), 2.38 (1H, br d, 8-H_{eq}), 2.40 (3H, s, 13-Me), 2.95 (1H, d, $J_{10\text{ax},10\text{eq}}=19$ Hz, 10-H_{ax}), 3.01 (1H, dd, $J_{3',3'\text{-OH}}=11.5$, $J_{3'\text{-OH},3'\text{-F}}=6$ Hz, 3'-OH), 3.21 (1H, dd, $J_{8\text{eq},10\text{eq}}\sim 2$ Hz, 10-H_{eq}), 3.44 (1H, ddd, $J=12$, 9, and 2.5 Hz, 6''-H_a), 3.74 (1H, m, 3'-H; collapsed to dt on deuteration, $J_{2',3'}=J_{3',4'}\sim 3$ Hz), 3.79 (1H, m, 4'-H), 3.94 (1H, m, 6''-H_b), 4.07 (1H, s, 9-OH), 4.09 (3H, s, OMe), 4.17 (1H, br q, 5'-H), 4.50 (1H, br d, $J_{2',2'\text{-F}}\sim 50$ Hz, 2'-H), 5.31 (1H, dd, $J_{7,8\text{eq}}\sim 1.5$ Hz, 7-H), 5.64 (1H, dd, $J_{1',2'\text{-F}}=10$, $J_{1',2'}=1.5$ Hz, 1'-H), 7.40 (1H, dd, $J_{2,3}=8.5$, $J_{1,3}\sim 1$ Hz, 3-H), 7.78 (1H, t, 2-H), 8.03 (1H, dd, $J_{1,2}=7.5$ Hz, 1-H), 13.25 and 14.00 (each 1H, s, 6, 11-OH); ¹⁹F NMR (CDCl_3) δ -204.8 (1F, ddd, $J=49$, 31 (= $J_{3',2'\text{-F}}$), and 10 Hz, 2'-F).

Anal Calcd for $\text{C}_{32}\text{H}_{34}\text{F}_2\text{O}_{12}$: C 59.26, H 5.28, F 5.86.

Found: C 59.34, H 5.41, F 5.70.

7b: ¹H NMR (CDCl_3) δ 1.41 (1H, d, $J_{5',6'}=6.5$ Hz, 5'-Me), 1.63~1.84 (2H, m, 5''-H_a, H_b), ~1.94 (1H, m, 4''-H_a), ~2.1 (1H, m, 4''-H_b), 2.17 (1H, dd, $J_{7,8\text{ax}}=4.5$, $J_{8\text{ax},8\text{eq}}=15$ Hz, 8-H_{ax}), 2.37 (1H, dt, $J_{7,8\text{eq}}=J_{8\text{eq},10\text{eq}}\sim 2$ Hz, 8-H_{eq}), 2.40 (3H, s, 13-Me), 2.97 (1H, d, $J_{10\text{ax},10\text{eq}}=19$ Hz, 10-H_{ax}), 3.24 (1H, dd, 10-H_{eq}), 3.52~3.81 (3H, m, 3', 4'-H, 6''-H_a; 3'-H at δ 3.63 collapsed to dt on deuteration, $J_{3',2'\text{-F}}=33.5$, $J_{2',3'}=J_{3',4'}\sim 3$ Hz), 4.04 (1H, dd, $J=11.5$ and 4 Hz, 6''-H_b), 4.08 (1H, s, 9-OH), 4.09 (3H, s, OMe), 4.22 (1H, br q, 5'-H), 4.46 (1H, d, $J_{3',3'\text{-OH}}=11$ Hz, 3'-OH), 4.51 (1H, br d, $J_{2',2'\text{-F}}\sim 50$ Hz, 2'-H), 5.33 (1H, dd, 7-H), 5.67 (1H, br d, $J_{1',2'\text{-F}}=10$ Hz, 1'-H), 7.41 (1H, dd, $J_{2,3}=8.5$, $J_{1,3}\sim 1$ Hz, 3-H), 7.79 (1H, t, 2-H), 8.04 (1H, dd, $J_{1,2}=7.5$ Hz, 1-H), 13.27 and 14.02 (each 1H, s, 6, 11-OH); ¹⁹F NMR (CDCl_3) δ -203.0 (1F, ddd, $J=49$, 33.5, and 10 Hz, 2'-F).

Anal Calcd for $\text{C}_{32}\text{H}_{34}\text{F}_2\text{O}_{12}\cdot\frac{1}{2}\text{H}_2\text{O}$: C 58.44, H 5.36, F 5.78.

Found: C 58.70, H 5.39, F 5.64.

7c: ¹H NMR (CDCl_3) δ 1.38 (1H, d, $J_{5',6'}=6.5$ Hz, 5'-Me), 1.53~1.91 (3H, m, 4''-H_a, 5''-H_a, H_b), ~2.15 (1H, m, 4''-H_b), 2.17 (1H, dd, $J_{7,8\text{ax}}=4.5$, $J_{8\text{ax},8\text{eq}}=15$ Hz, 8-H_{ax}), 2.36 (1H, dt, $J_{7,8\text{eq}}=J_{8\text{eq},10\text{eq}}\sim 2$ Hz, 8-H_{eq}), 2.40 (3H, s, 13-Me), 2.95 (1H, d, $J_{10\text{ax},10\text{eq}}=19$ Hz, 10-H_{ax}), 3.23 (1H, dd, 10-H_{eq}), 3.57 (1H, ddd, $J\sim 11$, ~8, and ~3 Hz, 6''-H_a), 3.63 (1H, ddt, $J_{2',3'}=J_{3',4'}\sim 3$, $J_{3'\text{-OH}}=11$, $J_{3',2'\text{-F}}=33$ Hz, 3'-H; collapsed to dt on deuteration), 3.68 (1H, m, 4'-H), ~4.05 (1H, m, 6''-H_b), 4.07 (1H, s, 9-OH), 4.09 (3H, s, OMe), 4.21 (1H, br q, 5'-H), 4.46 (1H, d, 3'-OH), 4.49 (1H, br d, $J_{2',2'\text{-F}}\sim 50$ Hz, 2'-H), 5.33 (1H, dd, 7-H), 5.65 (1H, dd, $J_{1',2'}\sim 1.5$, $J_{1',2'\text{-F}}=10$ Hz, 1'-H), 7.40 (1H, dd, $J_{1,3}\sim 1$, $J_{2,3}=8.5$ Hz, 3-H), 7.78 (1H, t, 2-H), 8.03 (1H, dd, $J_{1,2}=7.5$ Hz, 1-H), 13.26 and 14.00 (each 1H, s, 6, 11-OH); ¹⁹F NMR (CDCl_3) δ -203.3 (1F, ddd, $J=49$, 33.5, and 10 Hz, 2'-F).

Anal Calcd for $\text{C}_{32}\text{H}_{34}\text{F}_2\text{O}_{12}$: C 59.26, H 5.28, F 5.86.

Found: C 59.06, H 5.30, F 5.87.

Acknowledgment

The authors wish to thank Central Research Laboratories of Meiji Seika Kaisha, Ltd. for the biological assays in Table 4.

References

- 1) TSUCHIYA, T.; Y. TAKAGI, K. OK, S. UMEZAWA, T. TAKEUCHI, N. WAKO & H. UMEZAWA: Syntheses and antitumor

- activities of 7-*O*-(2,6-dideoxy-2-fluoro- α -L-talopyranosyl)daunomycinone and -adriamycinone. *J. Antibiotics* 39: 731~733, 1986
- 2) OK, K.; Y. TAKAGI, T. TSUCHIYA, S. UMEZAWA & H. UMEZAWA: Synthesis of antitumor-active 7-*O*-(2,6-dideoxy-2-fluoro- α -L-talopyranosyl)-daunomycinone and -adriamycinone. *Carbohydr. Res.* 169: 69~81, 1987
 - 3) TSUCHIYA, T.; Y. TAKAGI, S. UMEZAWA, T. TAKEUCHI, K. KOMURO, C. NOSAKA, H. UMEZAWA, S. FUKATSU & T. YONETA: Synthesis and antitumor activities of 14-*O*-acyl derivatives of 7-*O*-(2,6-dideoxy-2-fluoro- α -L-talopyranosyl)adriamycinone. *J. Antibiotics* 41: 988~991, 1988
 - 4) TAKAGI, Y.; H. PARK, T. TSUCHIYA, S. UMEZAWA, T. TAKEUCHI, K. KOMURO & C. NOSAKA: Syntheses and antitumor activities of 7-*O*-(3-amino-2,3,6-trideoxy-2-fluoro- α -L-talopyranosyl)daunomycinone and -adriamycinone. *J. Antibiotics* 42: 1315~1317, 1989
 - 5) TAKAGI, Y.; N. KOBAYASHI, T. TSUCHIYA, S. UMEZAWA, T. TAKEUCHI, K. KOMURO & C. NOSAKA: Syntheses and antitumor activities of 7-*O*-(6-deoxy-2-*O*-methyl- α -L-talopyranosyl)daunomycinone and -adriamycinone. *J. Antibiotics* 42: 1318~1320, 1989
 - 6) OKI, T.; Y. MATSUZAWA, A. YOSHIMOTO, K. NUMATA, I. KITAMURA, S. HORI, A. TAKAMATSU, H. UMEZAWA, M. ISHIZUKA, H. NAGANAWA, H. SUDA, M. HAMADA & T. TAKEUCHI: New antitumor antibiotics, aclacinomycins A and B. *J. Antibiotics* 28: 830~834, 1975
 - 7) OKI, T.; I. KITAMURA, Y. MATSUZAWA, N. SHIBAMOTO, T. OGASAWARA, A. YOSHIMOTO, T. INUI, H. NAGANAWA, T. TAKEUCHI & H. UMEZAWA: Antitumor anthracycline antibiotics, aclacinomycin A and analogues. II. Structural determination. *J. Antibiotics* 32: 801~819, 1979
 - 8) UMEZAWA, H.; Y. TAKAHASHI, M. KINOSHITA, H. NAGANAWA, T. MASUDA, M. ISHIZUKA, K. TATSUTA & T. TAKEUCHI: Tetrahydropyranyl derivatives of daunomycin and adriamycin. *J. Antibiotics* 32: 1082~1084, 1979
 - 9) UMEZAWA, H.; Y. TAKAHASHI, T. TAKEUCHI, H. NAKAMURA, Y. IITAKA & K. TATSUTA: The absolute structures of THP-adriamycins. *J. Antibiotics* 37: 1094~1097, 1984
 - 10) DANTCHEV, D.; M. PAINTRAND, M. HAYAT, C. BOURUT & G. MATHÉ: Low heart and skin toxicity of a tetrahydropyranyl derivative of adriamycin (THP-ADM) as observed by electron and light microscopy. *J. Antibiotics* 32: 1085~1086, 1979
 - 11) MATSUSHITA, Y.; H. KUMAGAI, A. YOSHIMOTO, H. TONE, T. ISHIKURA, T. TAKEUCHI & H. UMEZAWA: Antitumor activities of (2''*R*)-4'-*O*-tetrahydropyranyladriamycin (THP) and its combination with other antitumor agents on murine tumors. *J. Antibiotics* 38: 1408~1419, 1985
 - 12) KUNIMOTO, S.; K. MIURA, Y. TAKAHASHI, T. TAKEUCHI & H. UMEZAWA: Rapid uptake by cultured tumor cells and intracellular behavior of 4'-*O*-tetrahydropyranyladriamycin. *J. Antibiotics* 36: 312~317, 1983
 - 13) CSUK, R. & B. I. GLÄNZER: N.M.R. spectroscopy of fluorinated monosaccharides. *Adv. Carbohydr. Chem. Biochem.* 46: 73~177, 1988